



Mouse UCHL1 ELISA Kit

User's Manual

For Research Use Only, Not for use in diagnostic procedures

ELISA Kit for Measuring Mouse UCHL1

CircuLex Mouse UCHL1 ELISA Kit

Cat# CY-8093

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Intended Use

The CycLex Research Product **CircuLex Mouse UCHL1 ELISA Kit** is used for the quantitative measurement of mouse UCHL1 in cell lysate, cell culture supernatant and other biological media.

This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.

Storage

- Upon receipt store all components at 4°C.
- Don't expose reagents to excessive light.

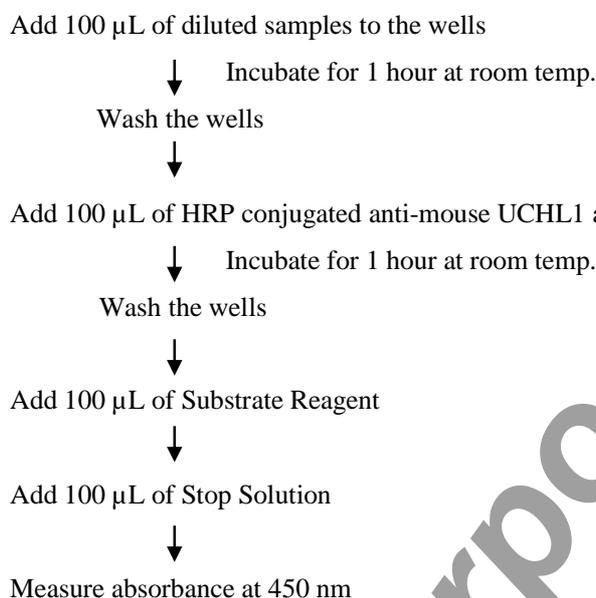
Introduction

The UCHL1, also called neuronal-specific protein gene product 9.5, is a carboxyl-terminal ubiquitin hydrolase regulating ubiquitin dependent signaling pathways, recently suggested as a tumor suppressor. UCHL1 is expressed predominantly in neurons (1), representing 1 to 2% of total soluble brain protein (2), as well as in testis and ovary. In vivo, UCHL1 has been shown to be involved in the regulation of the ubiquitin pool, apoptosis, learning and memory, and its absence in mice because of spontaneous intragenic deletions yields phenotypes with neurological defects (3). Mutations in UCHL1 have been discovered in a German family with Parkinson disease (PD); a point mutation near the active site that changes Ile 93 to Met (I93M), which caused a partial loss of the catalytic activity of this thiol protease. This mutation has been linked to an increased risk of developing an autosomal-dominant form of PD (4). Based on the abundance in the CNS, UCHL1 has been proposed as a candidate biomarker for brain injury and ischemic strokes. It was demonstrated that UCHL1 was released from injured neurons and flow into the cerebrospinal fluid and eventually into circulating blood (5).

Principle of the Assay

The CycLex Research Product **CircuLex Mouse UCHL1 ELISA Kit** employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for mouse UCHL1 is pre-coated onto a microplate. Standards and samples are pipetted into the wells and the immobilized antibody binds any mouse UCHL1 present. After washing away any unbound substances, an HRP conjugated antibody specific for mouse UCHL1 is added to the wells. Following a wash to remove any unbound antibody HRP conjugate, the remaining conjugate is allowed to react with the substrate H₂O₂-tetramethylbenzidine. The reaction is stopped by addition of acidic solution and absorbance of the resulting yellow product is measured at 450 nm. The absorbance is proportional to the concentration of Mouse UCHL1. A standard curve is constructed by plotting absorbance values versus mouse UCHL1 concentrations of calibrators, and concentrations of unknown samples are determined using this standard curve.

Summary of Procedure



Materials Provided

All samples and standards should be assayed in duplicate. The following components are supplied and are sufficient for the one 96-well microplate kit.

Microplate: One microplate supplied ready to use, with 96 wells (12 strips of 8-wells) in a foil, zip-lock bag with a desiccant pack. Wells are coated with anti-mouse UCHL1 antibody as a capture antibody.

10X Wash Buffer: One bottle containing 100 mL of 10X buffer containing 2% Tween®-20

Dilution Buffer: One bottle containing 50 mL of 1X buffer; use for reconstitution of Mouse UCHL1 Standard and sample dilution. Ready to use.

Mouse UCHL1 Standard: One vial containing 100 ng of lyophilized recombinant Mouse UCHL1.

HRP conjugated Detection Antibody: One bottle containing 12 mL of HRP (horseradish peroxidase) conjugated anti-mouse UCHL1 antibody. Ready to use.

Substrate Reagent: One bottle containing 20 mL of the chromogenic substrate, tetra-methylbenzidine (TMB). Ready to use.

Stop Solution: One bottle containing 20 mL of 1 N H₂SO₄. Ready to use.

Materials Required but not Provided

- **Pipettors:** 2-20 µL, 20-200 µL and 200-1000 µL precision pipettors with disposable tips
- **Precision repeating pipettor**
- **Orbital microplate shaker**
- **Microcentrifuge and tubes** for sample preparation
- **Vortex mixer**
- **Microplate washer:** optional (Manual washing is possible but not preferable)
- **Plate reader:** capable of measuring absorbance in 96-well plates at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. The plate can also be read at a single wavelength of 450 nm, which will give a somewhat higher reading.
- **Software package facilitating data generation and analysis :**optional
- **500 or 1000 mL graduated cylinder**
- **Reagent reservoirs**
- **Deionized water of the highest quality**
- **Disposable paper towels**

Precautions and Recommendations

- Allow all the components to come to room temperature before use.
- All microplate strips that are not immediately required should be returned to the zip-lock pouch, which must be carefully resealed to avoid moisture absorption.
- Do not use kit components beyond the indicated kit expiration date.
- Use only the microtiter wells provided with the kit.
- Rinse all detergent residues from glassware.
- Use deionized water of the highest quality.
- Do not mix reagents from different kits.
- The buffers and reagents in this kit may contain preservatives or other chemicals. Care should be taken to avoid direct contact with these reagents.
- Do not mouth pipette or ingest any of the reagents.
- Do not smoke, eat, or drink when performing the assay or in areas where samples or reagents are handled.
- Dispose of tetra-methylbenzidine (TMB) containing solutions in compliance with local regulations.
- Avoid contact with the acidic Stop Solution and Substrate Solution, which contains hydrogen peroxide.
- Wear gloves and eye protection when handling immunodiagnostic materials and samples of human origin, and these reagents. In case of contact with the Stop Solution and the Substrate Solution, wash skin thoroughly with water and seek medical attention, when necessary.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**
- **CAUTION: Sulfuric Acid is a strong acid. Wear disposable gloves and eye protection when handling Stop Solution.**

Sample Collection and Storage

Cell lysate: Prepare cell lysates. Assay immediately or store the samples on ice for a few hours before assaying. Aliquots of the samples may also be stored at below -70°C for extended periods of time. Avoid repeated freeze-thaw cycles. (See below.)

Cell culture supernatant: Remove any particulates by centrifugation and assay immediately or aliquot and store samples at below -70°C . Avoid repeated freeze-thaw cycles.

Other biological samples: Remove any particulates by centrifugation and assay immediately or aliquot and store samples at below -70°C . Avoid repeated freeze-thaw cycles.

Detailed Protocol

The CycLex Research Product **CircuLex Mouse UCHL1 ELISA Kit** is provided with removable strips of wells so the assay can be carried out on separate occasions using only the number of strips required for the particular determination. Since experimental conditions may vary, an aliquot of the mouse UCHL1 Standard within the kit should be included in each assay as a calibrator. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

Preparation of Working Solutions

All reagents need to be brought to room temperature prior to the assay. Assay reagents are supplied ready-to-use, with the exception of **10X Wash Buffer** and **Mouse UCHL1 Standard**.

1. Prepare a working solution of Wash Buffer by adding 100 mL of the 10X Wash Buffer (provided) to 900 mL of deionized (distilled) water. Mix well. Store at 4°C for two weeks or -20°C for long-term storage.
2. Reconstitute **Mouse UCHL1 Standard** with **1.0 mL** of **Dilution Buffer**. The concentration of the mouse UCHL1 in vial should be **100 ng/mL**, which is referred as a **Master Standard** of mouse UCHL1.

Prepare Standard solutions as follows:

Use the **Master Standard** to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 20 ng/mL standard (Std.1) serves as the highest standard. The **Dilution Buffer** serves as the zero standard (Blank).

	Volume of Standard	Dilution Buffer	Concentration
Std.1	120 µL of Master Standard (200 ng/mL)	480 µL	20 ng/mL
Std.2	300 µL of Std. 1 (20 ng/mL)	300 µL	10 ng/mL
Std.3	300 µL of Std. 2 (10 ng/mL)	300 µL	5 ng/mL
Std.4	300 µL of Std. 3 (5 ng/mL)	300 µL	2.5 ng/mL
Std.5	300 µL of Std. 4 (2.5 ng/mL)	300 µL	1.25 ng/mL
Std.6	300 µL of Std. 5 (1.25 ng/mL)	300 µL	0.63 ng/mL
Std.7	300 µL of Std. 6 (0.625 ng/mL)	300 µL	0.31 ng/mL
Blank		300 µL	0 ng/mL

Note: Do not use a Repeating pipette. Change tips for every dilution. Wet tip with Dilution Buffer before dispensing. Unused portions of Master Standard should be aliquoted and stored at below -70°C immediately. Avoid multiple freeze and thaw cycles.

Sample Preparation

- Cell lysates and other biological samples might be required 10- to 100-fold dilution.
- Cell culture supernatants may not require dilution.

Standard Assay Procedure for Mouse UCHL1

1. Remove the appropriate number of microtiter wells from the foil pouch and place them into the well holder. Return any unused wells to the foil pouch, refold, seal with tape and store at 4°C.
2. Dilute samples with Dilution Buffer. (See "Sample Preparation" above.)
3. Pipette **100 µL** of **Mouse UCHL1 Standards (Std1-Std7, Blank)** and **diluted samples** in duplicates, into the appropriate wells.
4. Incubate the plate at room temperature (ca.25°C) for 1 hour, shaking at ca. 300 rpm on an orbital microplate shaker.
5. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
6. Add **100 µL** of **HRP conjugated Detection Antibody** into each well.
7. Incubate the plate at room temperature (ca.25°C) for 1 hour, shaking at ca. 300 rpm on an orbital microplate shaker.
8. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
9. Add **100 µL** of **Substrate Reagent**. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with e.g. aluminum foil is recommended. Return Substrate Reagent to 4°C immediately after the necessary volume is removed
10. Incubate the plate at room temperature (ca.25°C) for 10-20 minutes, shaking at ca. 300 rpm on an orbital microplate shaker. The incubation time may be extended up to 30 minutes if the reaction temperature is below than 20°C.
11. Add **100 µL** of **Stop Solution** to each well in the same order as the previously added Substrate Reagent.
12. Measure absorbance in each well using a spectrophotometric microplate reader at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. Read the microplate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop Solution.

Note-1: Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

Note-2: Reliable standard curves are obtained when either O.D. values do not exceed 0.25 units for the blank (zero concentration), or 3.0 units for the highest standard concentration. The plate should be monitored at 5-minute intervals for approximately 30 minutes.

Note-3: If the microplate reader is not capable of reading absorbance greater than the absorbance of the highest standard, perform a second reading at 405 nm. A new standard curve, constructed using the values measured at 405 nm, is used to determine mouse UCHL1 concentration of off-scale samples. The readings at 405 nm should not replace the on-scale readings at 450 nm.

Calculations

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Plot the optical density for the standards versus the concentration of the standards and draw the best curve. The data can be linearized by using log/log paper and regression analysis may be applied to the log transformation. To determine the mouse UCHL1 concentration of each sample, first find the absorbance value on the y-axis and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the x-axis and read the corresponding mouse UCHL1 concentration. If the samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

1. The dose-response curve of this assay fits best to a sigmoidal 4-parameter logistic equation. The results of unknown samples can be calculated with any computer program having a 4-parameter logistic function. It is important to make an appropriate mathematical adjustment to accommodate for the dilution factor.
2. Most microtiter plate readers perform automatic calculations of analyte concentration. The calibration curve is constructed by plotting the absorbance (Y) of calibrators versus log of the known concentration (X) of calibrators, using the 4-parameter function. Alternatively, the logit log function can be used to linearize the calibration curve (i.e. logit of absorbance (Y) is plotted versus log of the known concentration (X) of calibrators).

Measurement Range

The measurement range is 0.31 ng/mL to 20 ng/mL. Any sample reading higher than the highest standard should be diluted with Dilution Buffer in higher dilution and re-assayed. Dilution factors need to be taken into consideration in calculating the mouse UCHL1 concentration.

Troubleshooting

1. The Mouse UCHL1 Standard should be run in duplicate, using the protocol described in the **Detailed Protocol**. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. Poor duplicates, accompanied by elevated values for wells containing no sample, indicate insufficient washing. If all instructions in the **Detailed Protocol** were followed accurately, such results indicate a need for washer maintenance.
3. Overall low signal may indicate that desiccation of the plate has occurred between the final wash and addition of Substrate Reagent. Do not allow the plate to dry out. Add Substrate Reagent immediately after wash.

Reagent Stability

All of the reagents included in the CycLex Research Product **CircuLex Mouse UCHL1 ELISA Kit** have been tested for stability. Reagents should not be used beyond the stated expiration date. Upon receipt, kit reagents should be stored at 4°C, except the reconstituted UCHL1 Standard must be stored at below -70°C. Coated assay plates should be stored in the original foil bag sealed by the zip lock and containing a desiccant pack.

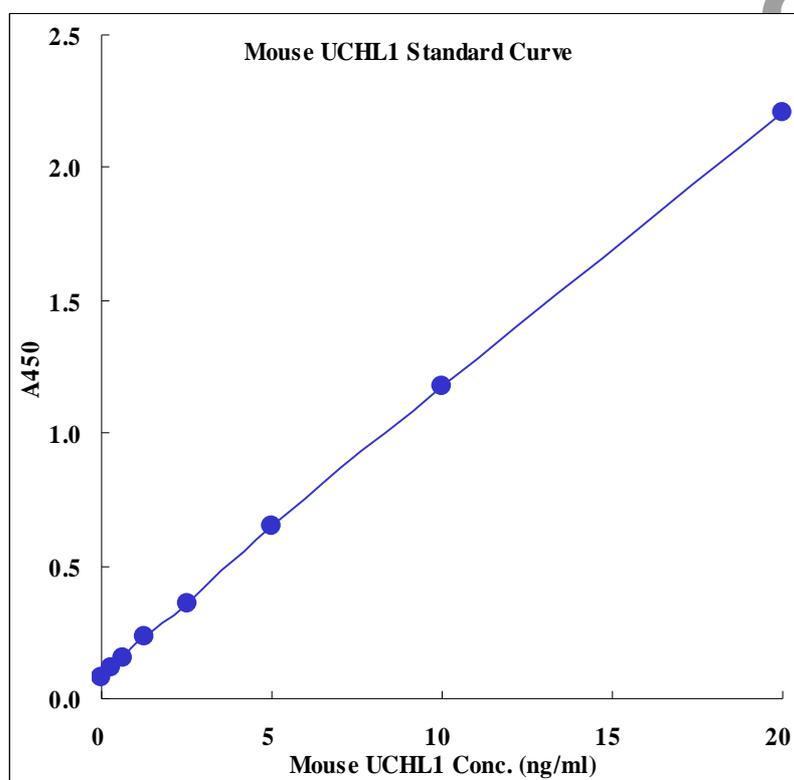
Assay Characteristics

1. Sensitivity

The limit of detection (defined as such a concentration of mouse UCHL1 giving absorbance higher than mean absorbance of blank* plus three standard deviations of the absorbance of blank: A blank + 3SD blank) is better than 229 pg/ml of sample.

* Dilution Buffer is pipetted into blank wells.

Typical Standard Curve



2. Precision

Intra-assay Precision (Precision within an assay)

A sample of known concentration was tested seven times on one plate to assess intra-assay precision.

- Intra-assay (Within-Run, n=7) CV=5.7 %

Mouse UCHL1 concentration (pg/mg total protein)

No.	Sample 1
1	796.8
2	705.4
3	724.1
4	790.5
5	801.3
6	713.8
7	724.3
max.	801.3
min.	705.4
mean	750.9
SD	43.0
CV(%)	5.7

Inter-assay Precision (Precision between assays)

A sample of known concentration was tested in three separate assays to assess inter-assay precision.

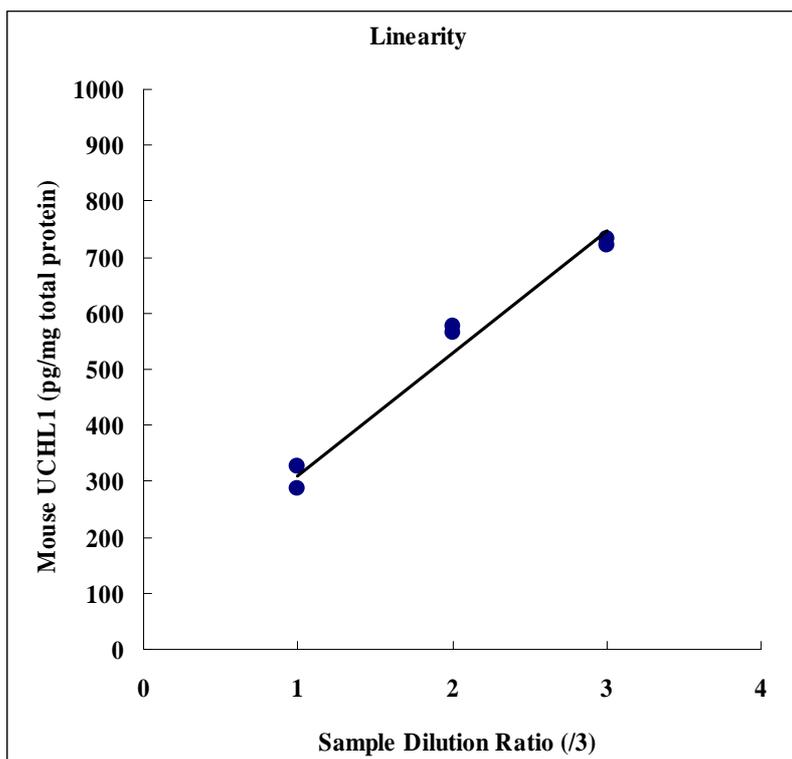
- Inter-assay (Run-to-Run, n=3) CV=0.7 %

Mouse UCHL1 concentration (pg/mg total protein)

Assay	Sample 1
1	764.2
2	754.2
3	758.0
max.	764.2
min.	754.2
mean	758.8
SD	5.0
CV(%)	0.7

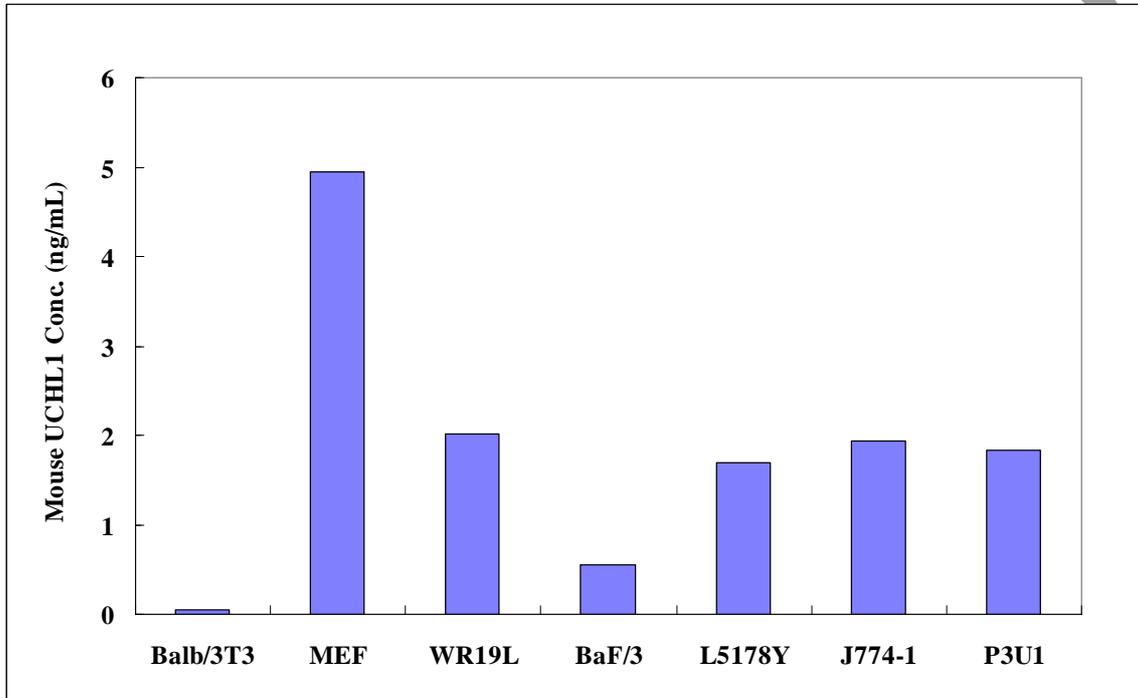
3. Linearity

One biological sample were diluted with Dilution Buffer and assayed after dilution. The neat sample is set to 1 (3/3). Please note a neat sample is 3-fold diluted as stated in the Assay Procedure. The result is summarized in the figure below.



Example of Test Results

Fig.1 Mouse UCHL1 concentrations in several cell culture supernatants



References

1. Osaka H, Wang YL, et al. (2003) Ubiquitin carboxy-terminal hydrolase L1 binds to and stabilizes monoubiquitin in neuron. Hum Mol Genet 12: 1945-58.
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3. Saigoh K, Wang YL, et al. (1999) Intragenic deletion in the gene encoding ubiquitin carboxy-terminal hydrolase in gad mice. Nat Genet 23: 47-51.
4. Leroy, E., Boyer, R. et al. (1998) The ubiquitin pathway in Parkinson's disease. Nature 395: 451-452.
5. Papa L, Akinyi L et al. (2010) Ubiquitin C-terminal hydrolase is a novel biomarker in humans for severe traumatic brain injury. Crit Care Med. 38: 138-44.

Related Products

- * CircuLex Human UCHL1 ELISA Kit: Cat# CY-8092
- * CircuLex Human DJ-1/PARK7 ELISA Kit: Cat# CY-9050
- * CircuLex Human 14-3-3 Gamma ELISA Kit: Cat# CY-8082
- * CycLex Poly-Ubiquitinated Protein ELISA Kit: Cat# CY-7053
- * CycLex Poly-Ubiquitinated Protein Enrichment & Detection Kit: Cat# CY-7001
- * CycLex Proteasone Enrichment & Activity Assay Kit: Cat# CY-7002

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