



Mouse Visfatin/PBEF ELISA Kit

User's Manual

For Research Use Only, Not for use in diagnostic procedures

ELISA Kit for Measuring Mouse Visfatin/PBEF

CircuLex Mouse Visfatin/PBEF ELISA Kit

Cat# CY-8065

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Intended Use

The CycLex Research Product **Circulex Mouse Visfatin/PBEF ELISA Kit** is used for the quantitative measurement of Mouse Visfatin/PBEF in serum, plasma, tissue culture medium and other biological media.

This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.

Storage

- Upon receipt store all components at 4°C.
- Don't expose reagents to excessive light.

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Introduction

Human obesity-related diabetes and the accompanying metabolic disorders have been specifically linked to increased visceral adipose tissue mass. Recently, visfatin was identified as a adipokine predominantly expressed in and secreted from visceral adipose tissue in both humans and mice by a differential display method using paired samples of visceral and subcutaneous fat from only two female volunteers (1). This adipokine was previously described as a growth factor for early B-cells called pre-B-cell colony-enhancing factor (2). Visfatin activated the insulin receptor in various insulin-sensitive cell types in vitro and visfatin treatment of mice acutely lowered plasma glucose in vivo. Moreover, mice heterozygous for a loss-of-function mutation in the visfatin gene had higher plasma glucose levels compared with wild-type littermates. In humans, visceral fat mass estimated by computed tomography was strongly correlated with plasma visfatin levels, whereas only a weak relationship with subcutaneous fat was observed (1). These findings suggested that visfatin could play a role in the association between visceral obesity and increased metabolic risk. However, its pathophysiological role in humans remains largely unknown.

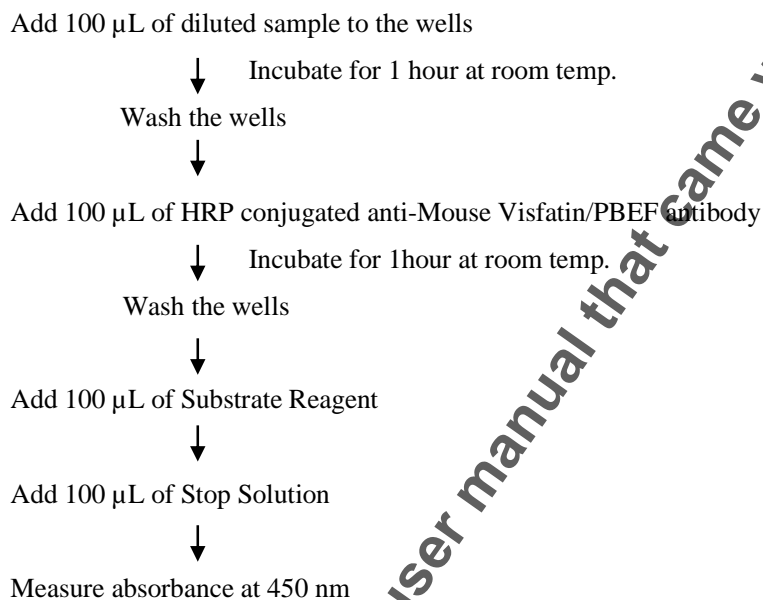
Principle of the Assay

The CircuLex Mouse Visfatin/PBEF ELISA Kit employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for Mouse Visfatin/PBEF has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and the immobilized antibody binds any Mouse Visfatin/PBEF present. After washing away any unbound substances, an HRP conjugated antibody specific for Mouse Visfatin/PBEF is added to the wells. Following a wash to remove any unbound antibody HRP conjugate, the remaining conjugate is allowed to react with the substrate H₂O₂-tetramethylbenzidine. The reaction is stopped by addition of acidic solution and absorbance of the resulting yellow product is measured at 450 nm. The absorbance is proportional to the concentration of Mouse Visfatin/PBEF. A standard curve is constructed by plotting absorbance values versus Mouse Visfatin/PBEF concentrations of calibrators and concentrations of unknown samples are determined using this standard curve.

The CircuLex Mouse Visfatin/PBEF ELISA Kit is designed to measure the concentration of Mouse Visfatin/PBEF from mouse serum/plasma, cultured mouse macrophages, or conditioned medium.

For Reference Purpose Only! Please refer to the user manual that came with your product.

Summary of Procedure



For Reference Purpose Only! Please refer to the user manual that came with your product.

Materials Provided

All samples and standards should be assayed in duplicate. The following components are supplied and are sufficient for the one 96-well microplate kit.

Microplate: One microplate supplied ready to use, with 96 wells (12 strips of 8-wells) in a foil, zip-lock bag with a desiccant pack. Wells are coated with anti-Mouse Visfatin/PBEF antibody as a capture antibody.

10X Wash Buffer: One bottle containing 100 mL of 10X buffer containing Tween®-20

Dilution Buffer: One bottle containing 50 mL of 1X buffer; use for sample dilution. Ready to use.

Mouse Visfatin/PBEF Standard: One vial containing 0.84 µg of lyophilized recombinant Mouse Visfatin/PBEF

HRP conjugated Detection Antibody: One bottle containing 12 mL of HRP (horseradish peroxidase) conjugated anti-Mouse Visfatin/PBEF antibody. Ready to use.

Substrate Reagent: One bottle containing 20 mL of the chromogenic substrate, tetra-methylbenzidine (TMB). Ready to use.

Stop Solution: One bottle supplied ready to use, containing 20 mL of 1 N H₂SO₄.

Materials Required but not Provided

- **Pipettors:** 2-20 μ L, 20-200 μ L and 200-1000 μ L precision pipettors with disposable tips.
- **Precision repeating pipettor**
- **Orbital microplate shaker**
- **Microcentrifuge and tubes** for sample preparation.
- **Vortex mixer**
- **Microplate washer:** optional (Manual washing is possible but not preferable)
- **Plate reader** capable of measuring absorbance in 96-well plates at dual wavelengths of 450 nm/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. The plate can also be read at a single wavelength of 450 nm, which will give a somewhat higher reading.
- **Software package facilitating data generation and analysis:** optional
- **500 or 1000 mL graduated cylinder**
- **Reagent reservoirs**
- **Deionized water of the highest quality**
- **Disposable paper towels**

Precautions and Recommendations

- Allow all the components to come to room temperature before use.
- All microplate strips that are not immediately required should be returned to the zip-lock pouch, which must be carefully resealed to avoid moisture absorption.
- Do not use kit components beyond the indicated kit expiration date.
- Use only the microtiter wells provided with the kit.
- Rinse all detergent residue from glassware.
- Use deionized water of the highest quality.
- Do not mix reagents from different kits.
- The buffers and reagents in this kit may contain preservatives or other chemicals. Care should be taken to avoid direct contact with these reagents.
- Do not mouth pipette or ingest any of the reagents.
- Do not smoke, eat, or drink when performing the assay or in areas where samples or reagents are handled.
- Dispose of tetra-methylbenzidine (TMB) containing solutions in compliance with local regulations.
- Avoid contact with the acidic Stop Solution and Substrate Solution, which contains hydrogen peroxide.
- Wear gloves and eye protection when handling immunodiagnostic materials and samples of human origin, and these reagents. In case of contact with the Stop Solution and the Substrate Solution, wash skin thoroughly with water and seek medical attention, when necessary.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**
- **CAUTION: Sulfuric Acid is a strong acid. Wear disposable gloves and eye protection when handling Stop Solution.**

Sample Collection and Storage

Serum: Use a serum separator tube and allow samples to clot for 60 ± 30 minutes. Centrifuge the samples at 4°C for 10 minutes at 1,000 x g. Remove serum and assay immediately or store samples on ice for up to 6 hours before assaying. Aliquots of serum may also be stored at below -70°C for extended periods of time. Avoid repeated freeze-thaw cycles.

Plasma: Collect plasma using EDTA-Na₂ as the anticoagulant. If possible, collect the plasma into a mixture of EDTA-Na₂ and Futhan5 to stabilize the sample against spontaneous *in vitro* complement activation. Immediately centrifuge samples at 4°C for 15 minutes at 1,000 x g. Assay immediately or store samples on ice for up to 6 hours before assaying. Aliquots of plasma may also be stored at below -70°C for extended periods of time. Avoid repeated freeze-thaw cycles.

Note: Citrate plasma has not been validated for use in this assay.

Other biological samples: Remove any particulates by centrifugation and assay immediately or aliquot and store samples at below -70°C. Avoid repeated freeze-thaw cycles.

Detailed Protocol

The CycLex Research Product **CircuLex Mouse Visfatin/PBEF ELISA Kit** is provided with removable strips of wells so the assay can be carried out on separate occasions using only the number of strips required for the particular determination. Since experimental conditions may vary, an aliquot of the Mouse Visfatin/PBEF Standard within the kit, should be included in each assay as a calibrator. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

Preparation of Working Solutions

All reagents need to be brought to room temperature prior to the assay. Assay reagents are supplied ready-to-use, with the exception of **10X Wash Buffer** and **Mouse Visfatin/PBEF Standard**.

1. Prepare a working solution of Wash Buffer by adding 100 mL of the **10X Wash Buffer** to 900 mL of deionized (distilled) water. Mix well.
2. Reconstitute **Mouse Visfatin/PBEF Standard** with **0.7 mL** of **Dilution Buffer**. The concentration of the mouse Visfatin/PBEF in vial should be **1,200 ng/mL**, which is referred as a **Master Standard** of mouse Visfatin/PBEF.

Prepare Standard Solutions as follows:

Use the **Master Standard** to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 60 ng/mL standard (Std.1) serves as the highest standard. The **Dilution Buffer** serves as the zero standard (Blank).

	Volume of Standard	Dilution Buffer	Concentration
Std.1	50 µL of Master Standard	950 µL	60 ng/mL
Std.2	300 µL of Std. 1 (60 ng/ml)	300 µL	30 ng/mL
Std.3	300 µL of Std. 2 (30 ng/ml)	300 µL	15 ng/mL
Std.4	300 µL of Std. 3 (15 ng/ml)	300 µL	7.5 ng/mL
Std.5	300 µL of Std. 4 (7.5 ng/ml)	300 µL	3.75 ng/mL
Std.6	300 µL of Std. 5 (3.75 ng/ml)	300 µL	1.88 ng/mL
Std.7	300 µL of Std. 6 (1.88 ng/ml)	300 µL	0.94 ng/mL
Blank	-	300 µL	0 ng/mL

Note: Do not use a Repeating pipette. Change tips for every dilution. Wet tip with Dilution Buffer before dispensing. Unused portions of Standards should be aliquoted and stored at below -70°C immediately. Avoid multiple freeze and thaw cycles.

Sample Preparation

- Serum and plasma samples require a 10-fold dilution.
e.g. 25 µL sample + 225 µL Dilution Buffer

Assay Procedure

1. Remove the appropriate number of microtiter wells from the foil pouch and place them into the well holder. Return any unused wells to the foil pouch, refold, seal with tape and store at 4°C.
2. Dilute samples with Dilution Buffer. (See "Sample Preparation" above.)
3. Pipette **100 µL** of **Standard Solutions (Std1-Std7, Blank)** and **diluted samples** in duplicates, into the appropriate wells.
4. Incubate the plate at room temperature (ca.25°C) for 1 hour, shaking at ca. 300 rpm on an orbital microplate shaker.
5. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
6. Add **100 µL** of **HRP conjugated Detection Antibody** into each well.
7. Incubate the plate at room temperature (ca.25°C) for 1 hour, shaking at ca. 300 rpm on an orbital microplate shaker.
8. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
9. Add **100 µL** of **Substrate Reagent**. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with e.g. aluminum foil is recommended. Return Substrate Reagent to 4°C immediately after the necessary volume is removed
10. Incubate the plate at room temperature (ca. 25°C) for 10-20 minutes, shaking at ca. 300 rpm on an orbital microplate shaker. The incubation time may be extended up to 30 minutes if the reaction temperature is below than 20°C.
11. Add **100 µL** of **Stop Solution** to each well in the same order as the previously added Substrate Reagent.
12. Measure absorbance in each well using a spectrophotometric microplate reader at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. Read the microplate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop Solution*.

Note-1: Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

Note-2: Reliable standard curves are obtained when either O.D. values do not exceed 0.25 units for the blank (zero concentration), or 2.5 units for the highest standard concentration. The plate should be monitored at 5-minute intervals for approximately 30 minutes.

Note-3: If the microplate reader is not capable of reading absorbance greater than the absorbance of the highest standard, perform a second reading at 405 nm. A new standard curve, constructed using the values measured at 405 nm, is used to determine Mouse Visfatin/PBEF concentration of off-scale samples. The readings at 405 nm should not replace the on-scale

readings at 450 nm.

Calculations

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Plot the optical density for the standards versus the concentration of the standards and draw the best curve. The data can be linearized by using log/log paper and regression analysis may be applied to the log transformation. To determine the Mouse Visfatin/PBEF concentration of each sample, first find the absorbance value on the y-axis and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the x-axis and read the corresponding Mouse Visfatin/PBEF concentration. If the samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

1. The dose-response curve of this assay fits best to a sigmoidal 5-parameter logistic equation. The results of unknown samples can be calculated with any computer program having a 5-parameter logistic function. It is important to make an appropriate mathematical adjustment to accommodate for the dilution factor.
2. Most microtiter plate readers perform automatic calculations of analyte concentration. The calibration curve is constructed by plotting the absorbance (Y) of calibrators versus log of the known concentration (X) of calibrators, using the four-parameter function. Alternatively, the logit log function can be used to linearize the calibration curve (i.e. logit of absorbance (Y) is plotted versus log of the known concentration (X) of calibrators).

Measurement Range

The measurement range is 0.94 ng/mL to 60 ng/mL. Any sample reading higher than the highest standard should be diluted with Dilution Buffer in higher dilution and re-assayed. Dilution factors need to be taken into consideration in calculating the mouse Visfatin/PBEF concentration.

Troubleshooting

1. The Mouse Visfatin/PBEF Standard should be run in duplicate, using the protocol described in the **Detailed Protocol**. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. Poor duplicates, accompanied by elevated values for wells containing no sample, indicate insufficient washing. If all instructions in the **Detailed Protocol** were followed accurately, such results indicate a need for washer maintenance.
3. Overall low signal may indicate that desiccation of the plate has occurred between the final wash and addition of Substrate Reagent. Do not allow the plate to dry out. Add Substrate Reagent immediately after wash.

Reagent Stability

All of the reagents included in the CycLex Research Product **CircuLex Mouse Visfatin/PBEF ELISA Kit** have been tested for stability. Reagents should not be used beyond the stated expiration date. Upon receipt, kit reagents should be stored at 4°C, except the reconstituted Mouse Visfatin/PBEF Standard

must be stored at below -70°C. Coated assay plates should be stored in the original foil bag sealed by the zip lock and containing a desiccant pack.

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Assay Characteristics

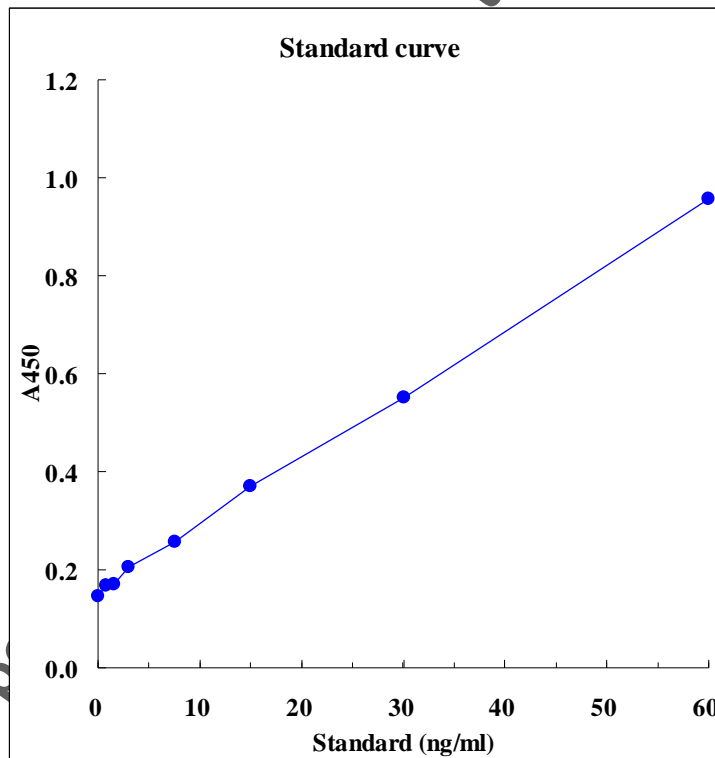
1. Sensitivity

The limit of detection (defined as such a concentration of Mouse Visfatin/PBEF giving absorbance higher than mean absorbance of blank* plus three standard deviations of the absorbance of blank: A blank + 3*SD blank) is better than 0.24 ng/ml of sample.

* Dilution Buffer is pipetted into blank wells.

Eighty assays were evaluated and the minimum detectable dose (MDD) of Mouse Visfatin/PBEF ranged from 0.179 - 0.291 ng/mL. The mean MDD was 0.243 ng/mL. The MDD was determined by adding three standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

Typical standard curve



2. Specificity

The antibodies in the CircuLex Mouse Visfatin/PBEF ELISA Kit are highly specific of Mouse Visfatin/PBEF, with detectable cross-reactivities to mouse Visfatin/PBEF and other cytokines that may be present in human serum.

3. Precision

Intra-assay Precision (Precision within an assay)

Three samples of known concentration were tested eight times on one plate to assess intra-assay precision.

- Intra-assay (Within-Run, n=8) CV=5.8 %

Inter-assay Precision (Precision between assays)

Three samples of known concentration were tested in four separate assays to assess inter-assay precision.

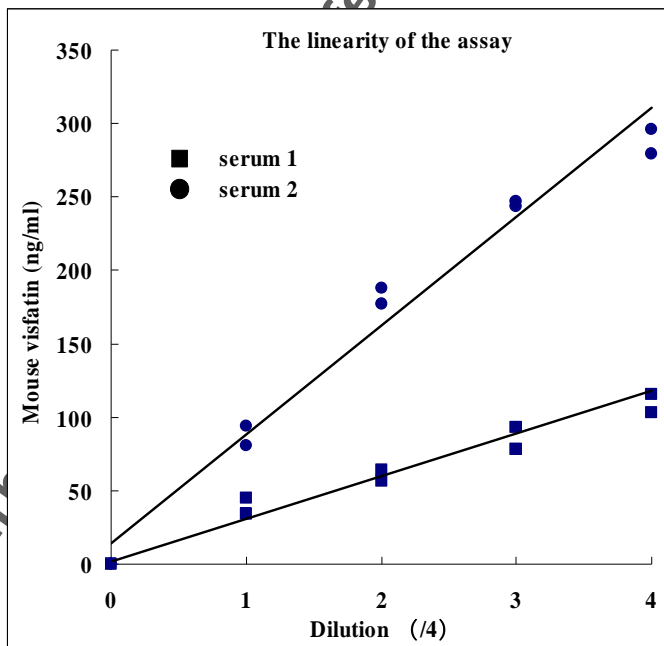
- Inter-assay (Run-to-Run, n=4) CV=8.2 %

4. Spiking Recover

Serum samples were spiked with different amounts of Mouse Visfatin/PBEF and assayed. The recovery of Mouse Visfatin/PBEF spiked to levels throughout the range of the assay was evaluated. Sample Average % Recovery Range
Cell culture media (n=4) 107, 88, 93, 114

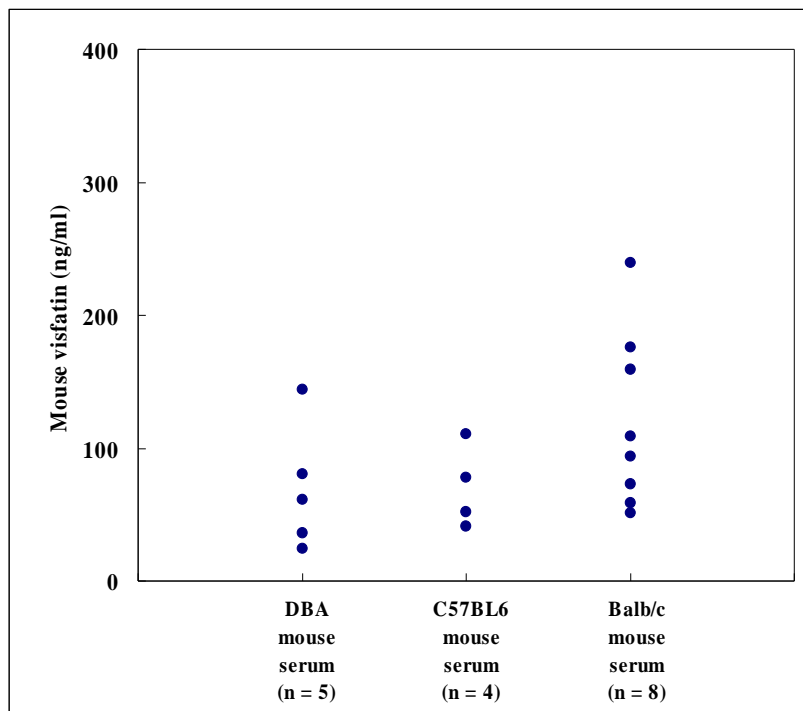
5. Linearity

To assess the linearity of the assay, samples containing and/or spiked with high concentrations of Mouse Visfatin/PBEF were serially diluted with the Dilution Buffer to produce samples with values within the dynamic range of the assay.



Example of Test Results

Fig.1 Concentrations of mouse visfatin in different strain mouse serum



References

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2. Samal B, Sun Y, Stearns G, Xie C, Suggs S, McNiece I: Cloning and characterization of the cDNA encoding a novel human pre-B-cell colony-enhancing factor. *Mol Cell Biol.* **14** :1431–1437, 1994
3. Miao-Pei Chen, Fu-Mei Chung, Dao-Ming Chang, Jack C.-R. Tsai, Han-Fen Huang, Shyi-Jang Shin, and Yau-Jiunn Lee: Elevated Plasma Level of Visfatin/Pre-B Cell Colony-Enhancing Factor in Patients with Type 2 Diabetes Mellitus *J. Clin. Endocrinol. Metab.* **91**, 295 – 299, 2006
4. Dominik G Haider, Karin Schindler, Georg Schaller, Gerhard Prager, Michael Wolzt, and Bernhard Ludvik: Increased plasma visfatin concentrations in morbidly obese subjects are reduced after gastric banding *J. Clin. Endocrinol. Metab.* **10**, 2005-2248, 2006

Related Products

- * CycLex NAMPT Colorimetric Assay Kit: Cat# CY-1251
- * NAMPT (Nicotinamide Phosphoribosyltransferase): Cat# CY-E1251
- * CircuLex Mouse Visfatin/PBEF ELISA Kit: Cat# CY-8065
- * Human NAMPT/Visfatin Low Endotoxin: Cat# CY-R2471
- * Anti-Human NAMPT Clone AF-1E12: Cat# CY-M1035
- * Anti-Human NAMPT: Cat# CY-P1038

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