



Chitotriosidase Fluorometric Assay Kit

User's Manual

For Research Use Only, Not for use in diagnostic procedures

Quantitative test kit for chitotriosidase activity

CycLex Chitotriosidase Fluorometric Assay Kit

For 100 Assays

Cat# CY-1249

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Intended Use

The CycLex Research Product **CycLex Chitotriosidase Fluorometric Assay Kit** is used for the quantitative measurement of chitotriosidase activity in serum/plasma, culture supernatant of activated macrophages and other biological samples.

Applications for this kit include:

- 1) Measuring chitotriosidase activity in serum/plasma and other biological samples.
- 2) Evaluating the effects of pharmacological compounds on chitotriosidase activity.
- 3) Screening inhibitors or activators of chitotriosidase.

This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.

Storage

- Upon receipt store Recombinant Chitotriosidase at -70°C and all other components below -20°C.
- Don't expose reagents to excessive light.



Introduction

Chitotriosidase was discovered in plasma of patients suffering from Gaucher's disease; it was found that the 1,000-fold-elevated enzyme originates from lipid-laden macrophages that accumulate in various tissues of Gaucher's patients (1). Chitotriosidase has subsequently been purified from the spleen of a Gaucher's patient and its cDNA was cloned from a human macrophage cDNA library (2, 3). This enzyme is a human chitinase member of family 18 glycosyl hydrolases (3–5), and has the capability to hydrolyze chitin. This enzyme is selectively expressed in activated tissue macrophages that accumulate in various tissues of several lysosomal diseases (6). Therefore its activity has been proposed as a biochemical marker of macrophage accumulation in Gaucher's disease (1, 7). In some other inherited lysosomal storage disorders, especially sphingolipidoses such as Niemann Pick, GM1-gangliosidosis, and Krabbe disease, which involve accumulation of different lipids, more modest elevations in plasma chitotriosidase have been noted (7). Chitotriosidase is the only biomarker identified up to date for the monitoring the efficacy of the extremely costly enzyme-replacement therapy of Gaucher patients and male Fabry patients (8).

Elevated levels of serum chitotriosidase were also found in disorders caused by the abnormal activation of immune system, including sarcoidosis (9) and atherosclerosis (10, 11). It has been shown that chitotriosidase activity was elevated up to 55-fold in extracts of atherosclerotic tissue, showing a clear connection between chitotriosidase expression and lipid-laden macrophages inside human atherosclerotic vessel wall (10). Human chitotriosidase also associates with pathogen-driven diseases, and in particular with fungal infections, suggesting the role of this enzyme in host defense against chitin-containing pathogens (12, 13). Other clinical data for instance show that chitotriosidase activity is raised in plasma of African children infected with acute *Plasmodium falciparum* malaria (14). Additional evidence for a role of chitotriosidase during immunological responses is the observation that the enzyme is shortly and acutely up-regulated both at the level of mRNA and activity following stimulation with prolactin, IFN- γ , TNF α and LPS, but not with IL-10 (15, 16).

In the blood stream, tissue macrophages largely secrete newly synthesized 50-kDa chitotriosidase, but about one-third is directly routed to lysosomes and proteolytically processed to the 39-kDa unit that remains catalytically active (17). A common chitotriosidase gene polymorphism leads to a null allele and therefore a defective enzyme activity. In white populations, 30% to 40% of individuals are carriers of this abnormal chitotriosidase allele and approximately 6% are homozygous (1, 18).



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Principle of the Assay

The **CycLex Chitotriosidase Fluorometric Assay Kit** is based on an exclusive fluorescence substrate, 4-Methylumbelliferyl- β -D-N,N',N''-triacetylchitotriose. This homogenous assay kit is sensitive and convenient.

Summary of Procedure

Dispense 5 μ L of sample or Recombinant chitotriosidase in the well.



Add 95 μ L of 1X Fluoro-Substrate solution.



Measure velocity of fluorescence intensity with excitation at 340-380 nm and emission at 440-460 nm for 30-60 min at 30°C.

Materials Provided

Each kit contains

Materials	Quantity	Storage
①10X Chitotriosidase Assay Buffer	1.0 mL x 2	Below -20°C
②50X Fluoro-Substrate (1 mM 4 MU-chitotriose*)	220 μ L x 1	Below -20°C
③Recombinant Chitotriosidase ** (ca. 5 μ g/ml)	50 μ L x 1	-70°C
④4-Methylumbelliferone standard (100 μ M)	200 μ L x 1	Below -20°C
⑤Instruction manual	1	room temp.

* 4-Methylumbelliferyl- β -D-N,N',N''-triacetylchitotriose

** Recombinant human chitotriosidase expressed in HEK293 cells.



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Materials Required but not Provided

- **Allosamidin:** Allosamidin is available from 2A PharmaChem; cat#. 2A-300198 (CAS number 103782-08-7).
- **Microplate suitable for use with a fluorometric plate reader** (black microplates provide better signal to noise ratio)
- **Microplate reading fluorometer:** capable of excitation at a wavelength in the range 340-380 nm and detection of emitted light in the range 440-460 nm
- **Pipettors:** 2-20 μL , 20-200 μL and 200-1000 μL precision pipettors with disposable tips.
- **Multi-channel pipette**
- **Microplate shaker**
- **Deionized water of the highest quality**
- **Reagent reservoirs**
- **Stop Solution (Optional):** Add 23.6 ml of deionized water to 2 g of sodium carbonate (Cat#: S2127) and mix well until completely dissolved. Store the Stop Solution (c.a. 1 M Na_2CO_3) at room temperature.

Precautions and Recommendations

- Please avoid repeated freezing and thawing of the Recombinant chitotriosidase in this kit. There is a possibility that the enzyme activity may be inactivated. Aliquot to 10 μL and store at -70°C .
- Do not use kit components beyond the indicated kit expiration date.
- Rinse all detergent residue from glassware.
- Use deionized water of the highest quality (ddH₂O).
- Do not mix reagents from different kits.
- Do not mouth pipette or ingest any of the reagents.
- Do not smoke, eat, or drink when performing the assay or in areas where samples or reagents are handled.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**



Detailed Protocol

The CycLex Research Product **CycLex Chitotriosidase Fluorometric Assay Kit** is provided with concentrated reagents. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

Preparation of Reagents within the kit

Thaw the reagents at room temperature except ③ **Recombinant Chitotriosidase** and keep all reagents on ice until use. Use them only after they are completely thawed and mixed.

#1. Prepare **1X Assay Buffer** by diluting the ① **10X Assay Buffer** (provided) **1 : 10** in distilled (deionized) water. Mix well. *Unused buffer should be stored at -20°C.*

#2. Prepare **Reaction Buffer** according to the table below. Mix well. *Discard any unused Reaction Buffer.*

Reaction Buffer

Assay reagents	1 assay	8 assays	16 assays	32 assays	48 assays
① 10X Assay Buffer	10 µL	80 µL	160 µL	320 µL	480 µL
② 50X Fluoro-Substrate	2 µL	16 µL	32 µL	64 µL	96 µL
Distilled water	88 µL	704 µL	1408 µL	2816 µL	4224 µL
Total volume	100 µL	800 µL	1600 µL	3200 µL	4800 µL

#3. Prepare **X10 diluted Recombinant Chitotriosidase** by diluting the ③ **Recombinant Chitotriosidase** (provided) **1 : 10** in #1. **1X Assay Buffer**. Mix well. Put it on ice. *Discard any unused X10 diluted Recombinant Chitotriosidase after diluted.*

Assay reagents	1 assay	8 assays	16 assays	32 assays	48 assays
#1. 1X Assay Buffer	9 µL	45 µL	81 µL	153 µL	225 µL
③ Recombinant Chitotriosidase	1 µL	5 µL	9 µL	17 µL	25 µL
Total volume	10 µL	50 µL	90 µL	170 µL	250 µL



Chitotriosidase Activity Assay Procedure

Following Table.1 below, first, add “**Test Sample**” or “**#3. X10 diluted Recombinant Chitotriosidase**” or “**#1. 1X Assay Buffer**” to each well of a microplate.

Table 1: Reaction mixture

Assay Reagents	Test Assay	Positive Control	No enzyme Control
Test Sample*	5 μ L	-	-
#3. X10 diluted Recombinant Chitotriosidase **	-	5 μ L	-
#1. 1X Assay Buffer**	-	-	5 μ L
#2. Reaction Buffer	95 μ L	95 μ L	95 μ L

* Test sample: Human serum/plasma sample or culture supernatant of activated macrophages. If sample might contain high concentration of chitotriosidase such as serum/plasma from Gaucher's disease patients, samples should be diluted 25-50 times in #1. 1X Assay Buffer before incubation.

** See page 5, section “Preparation of Reagents within the kit”

2) Finally, initiate the reaction by adding **95 μ L** of “**#2. Reaction Buffer**” * to each well and mixing thoroughly.

* See page 5, section “Preparation of Reagents within the kit”

3) Read fluorescence intensity for 30-60 min or desired length of time at 2 to 5 minute intervals using a microplate fluorometer with excitation at 340-380 nm and emission at 440-460 nm at 30°C*.

* Any assay temperature from room temperature to 37°C may be used.

4) Measure and calculate the rate of reaction while the reaction velocity remains constant.

Alternatively

3)' After incubation at 30°C for 20-30 min, the reaction was stopped by addition of **100 μ L** of **Stop Solution***. Fluorescence was read on a microplate fluorometer with excitation at 340-380 nm and emission at 440-460 nm

* See page 4, section “Materials Required but not Provided”



Inhibitor Screening Assay Procedure

In order to estimate the inhibitory effect on chitotriosidase activity by test compounds correctly, it is necessary to conduct the control experiment of “**Vehicle Control**” at least once for every experiment and “**Inhibitor Control**” at least once for the first experiment, in addition to “**Test Assay**” as indicated in the Table.1 (below). When test chemicals cause an inhibitory effect on Chitotriosidase activity, the level of increase of fluorescence intensity is weakened as compared with “**Vehicle Control**”. The increase in fluorescence intensity is not observed in “**Inhibitor Control**”.

- 1) Following Table.1 below, first, add “**#3. X10 diluted Recombinant Chitotriosidase**” or “**#1. 1X Assay Buffer**” to microplate wells. Second, add “**Test Compound**” or “**Vehicle of Test Compound**” or “**10X allosamidin**” to each well of the microplate and mix well*.

**Optional: For best accuracy, it is advisable to pre-incubate the plate for 5-10 min. at assay temperature.*

Table 1: Reaction mixture

Assay Reagents	Test Assay	Vehicle Control	Inhibitor Control	No Enzyme Control
#3. X10 diluted Recombinant Chitotriosidase *	5 μ L	5 μ L	5 μ L	-
#1. 1X Assay Buffer *	-	-	-	5 μ L
Test Compound	5 μ L	-	-	-
Vehicle of Test Compound	-	5 μ L	-	5 μ L
10X allosamidin (500 μ M) **	-	-	5 μ L	-
#2. Reaction Buffer *	90 μ L	90 μ L	90 μ L	90 μ L

* See page 5, section “Preparation of Reagents within the kit”

** Not provided in this kit. See page 4, section “Materials Required but not Provided”

- 2) Initiate reactions by adding 90 μ L of “**#2. Reaction Buffer**” * to each well and mixing thoroughly.

** See page 5, section “Preparation of Reagents within the kit”*

- 3) Read fluorescence intensity for 30-60 min or desired length of time at 2 to 5 minute intervals using microtiter plate fluorometer with excitation at 340-380 nm and emission at 440-460 nm at 30°C*.

** Any assay temperature from room temperature to 37°C may be used.*

- 4) Measure and calculate the rate of reaction while the reaction velocity remains constant.

Caution and Significance

- All assays should be assayed in duplicate.
- Use of a microplate shaker is recommended for complete mixing.
- If the test compounds or samples themselves emit fluorescence at excitation wavelength: 340-380 nm and fluorescence wavelength: 440-460 nm, the test assay cannot be evaluated correctly.



Determine microplate reader conversion factor for 4-Methylumbelliferone

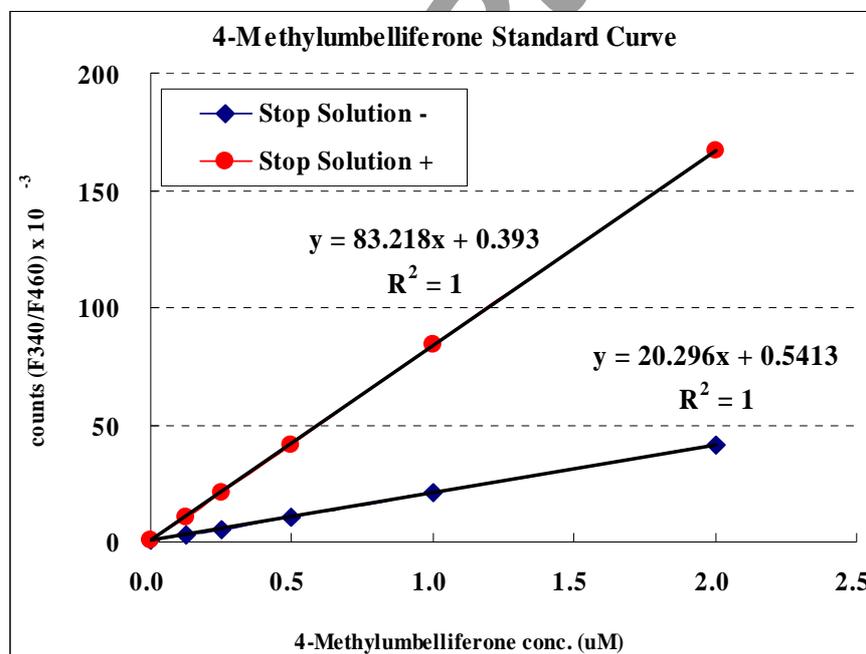
The exact 4-Methylumbelliferone concentration range that will be useful for preparing a standard curve will vary depending on the fluorometer model, the gain setting, and the exact excitation and emission wavelengths used. Please dilute the ④4-Methylumbelliferone standard (provided; 100 μM) to 1.0 μM as the highest standard and make a 4-fold serial dilution with Assay Buffer and then measuring the fluorescence of 100 μl in a microtiter plate fluorometer with excitation at 340-380 nm and emission at 440-460 nm. The estimate of $\mu\text{M}/\text{RFU}$ obtained with this measurement, together with the observed range of values obtained in the enzyme assays, can then be used to plan an appropriate series of dilutions for a standard curve. The slope of the standard curve can then be used as the $\mu\text{M}/\text{RFU}$ conversion factor.

When Stop Solution* is added to samples after enzyme reaction**, it also should be added to this 4-Methylumbelliferone standard curve. The addition of Stop Solution will give 4-Methylumbelliferone approximately a 4-fold higher fluorescence intensity than no addition.

* See page 4, section "Materials Required but not Provided"

** See page 6, "Alternatively" in the section "Chitotriosidase Activity Assay Procedure"

Typical 4-Methylumbelliferone Standard Curve





Evaluation of Results

Analysis of Kinetics

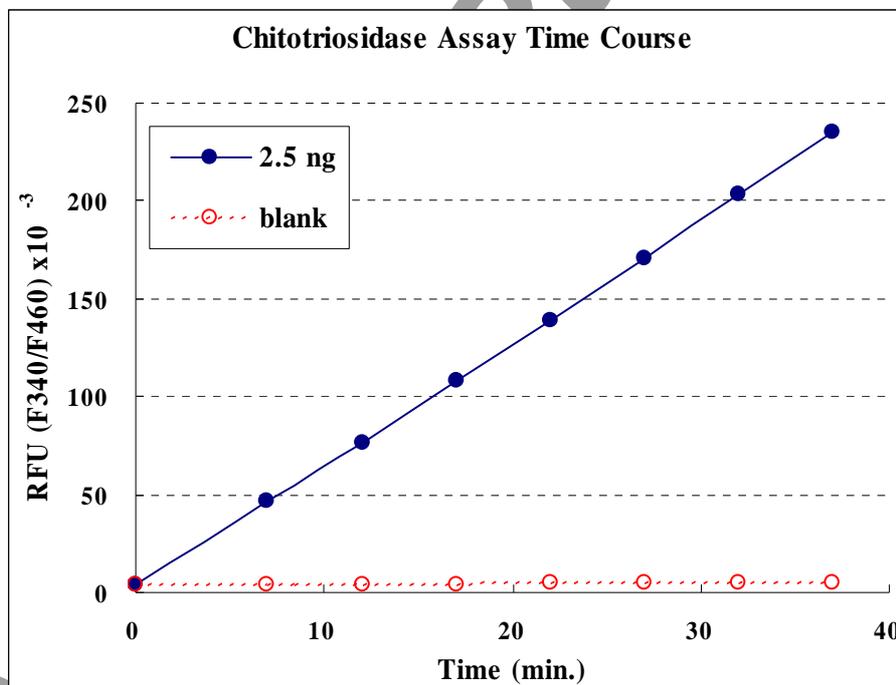
Time course curve

1. Run reactions as described in the *Detailed Protocol*.
2. Subtract fluorescence intensity at time 0 from all reaction time points.
3. Plot fluorescence intensity at 440-460 nm versus reaction time.
4. Determine the reaction time range in which the increase in fluorescence intensity at 440-460 nm is linear.
5. Calculate activity:

$$\text{Activity (reaction velocity)} = \frac{\text{Fluorescence Intensity of Test Assay}}{\text{Reaction time (min.)}}$$

NOTE: Usually, the linear range is from 0 to 30 min. This value is variable depending on reaction conditions and storage/handling of the Recombinant Chitotriosidase. Decreasing the amount of Recombinant Chitotriosidase in the assay may help to lengthen the time range.

Fig.1 Typical Time Course Curve





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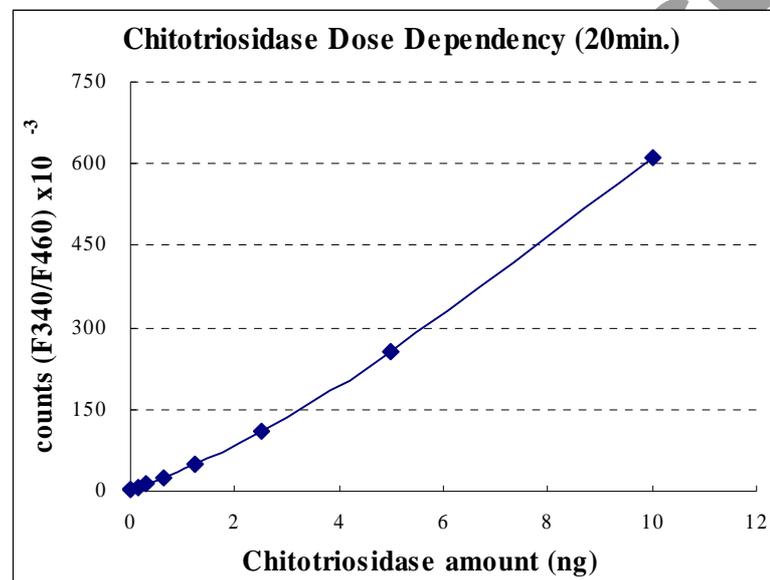
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Chitotriosidase Standard Curve and % Activity

1. Make serial dilutions of #3. X10 diluted Recombinant Chitotriosidase with #1. 1X Assay Buffer (ex. 100 %, 50 %, 25 %, 12.5 %, 6.25 %, 3.13 % and 0 %).
2. Run reactions with Vehicle and serial dilutions of Recombinant Chitotriosidase as described in the Detailed Protocol.
3. Plot standard curve data as fluorescence intensity at 460 nm versus dose of Chitotriosidase (ng/assay).
4. Obtain a line-fit to the data using appropriate calculations.
5. Use the slope and Y-intercept to calculate the amount of Chitotriosidase activity for the experimental data.

Fig.2 Typical Dose Dependency Curve



Analysis of Inhibitor Effect

% Intensity

1. Run reactions with test compounds and Vehicle as described in the **Detailed Protocol**.
2. Subtract fluorescence intensity of “No Enzyme Control” from all other experimental samples (Test Assay, Vehicle Control and Inhibitor Control).
3. Calculate the % Intensity:

$$\% \text{ Intensity} = \frac{\text{Reaction velocity of Test Assay}}{\text{Reaction velocity of Vehicle Control}} \times 100$$

NOTE: This % Intensity is a rough value of enzyme activity or inhibition. For greater accuracy, plot a standard curve of Chitotriosidase for each new set of reactions and estimate the % Activity (see above section “Chitotriosidase Standard Curve and % Activity”).



Troubleshooting

1. All assays should be run in duplicate using the protocol described in the *Detailed Protocol*. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. The reaction curve is nearly a straight line if the kinetics of the assay is of the first order. Variations in the protocol can lead to non-linearity of the curve, as can assay kinetics of other than first order. For a non-linear curve, point to point or quadratic curve fit methods should be used.
3. Poor duplicates, accompanied by elevated values for wells containing no sample, indicate inaccurate dispensing of assay reagents. If all instructions in the *Detailed Protocol* were followed accurately, such results indicate a need for multi-channel pipette maintenance.

Reagent Stability

All of the reagents included in the **CycLex Chitotriosidase Fluorometric Assay Kit** have been tested for stability. Reagents should not be used beyond the stated expiration date. Upon receipt, all kit reagents should be stored at -70°C . Avoid repeated freeze-thaw cycles of “**③Recombinant Chitotriosidase**”. After use, return kit reagents to -70°C as soon as possible.

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Related Products

*CircuLex Human Chitotriosidase ELISA Kit: Cat# CY-8074

*Human Chitotriosidase: Cat# CY-E1249

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