



FGFR2 Kinase Assay/Inhibitor Screening Kit

User's Manual

For Research Use Only, Not for use in diagnostic procedures

Non-Radioisotopic Kit for Measuring FGFR2 Kinase Activity

CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit

Cat# CY-1082

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Intended Use

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** is designed to measure the activities of recombinant catalytic domain of FGFR2 (Fibroblast Growth Factor Receptor 2) for the rapid and sensitive evaluation of inhibitors or activators. The phosphotyrosine specific monoclonal antibody used in this assay kit has been demonstrated to recognize the phosphotyrosine residue in recombinant “Tyrosine kinase-substrate-1”, which is efficiently phosphorylated by FGFR2 *in vitro*.

Applications of this kit include:

- 1) Screening inhibitors or activators of recombinant catalytic domain of FGFR2.
- 2) Detecting the effects of pharmacological agents on recombinant catalytic domain of FGFR2.

This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.

Storage

- Upon receipt store all components at 4°C.
- Don't expose reagents to excessive light.

For Reference Purpose Only! Please refer to the user manual that came with your product.



Introduction

The fibroblast growth factors (FGFs) belong to a family of about 23 related polypeptides. They display biological activity toward cells of mesenchymal, neuronal, and epithelial origin and are involved in processes such as cell growth, organ development, and angiogenesis (1). The biological effects of FGFs are exerted through interactions with FGF receptors (FGFRs). The receptor family consists of four known members, FGFR1-4, with many isoforms (2). Upon ligand binding the receptor is thought to be activated through dimerization and phosphorylation by the intracellular tyrosine kinase domains (3). Heparan sulfate proteoglycans (HSPGs), abundant components of cell surfaces and the extracellular matrix, appear central to signaling through FGF-FGFR complexes.

Overexpression and amplification of growth factor receptors are common alterations observed in human breast cancer cells. The FGFR1 and FGFR2 genes are expressed in both normal and breast cancer tissues (4), and alterations, including amplification and overexpression of FGFR1 and FGFR2, have previously been reported in 5–10% of primary breast cancer specimens (5, 6).

In this decade, because of the localization of FGFR2 to the same chromosomal region as the mutation responsible for Crouzon syndrome (7), FGFR2 became a candidate gene for the clinical disorder. It has been demonstrated that the mutations in FGFR2 gene was found in patients with Crouzon syndrome (8, 9), Apert syndrome (10), Pfeiffer syndrome (11, 12), and Jackson-Weiss syndrome (8, 13).

Measurement of FGFR2 Kinase activity

The protocol generally regarded as most sensitive for the quantitative measurement of FGFR2 kinase activity involves incubation of the FGFR2 kinase sample with substrate, either a natural or synthetic polypeptide (such as poly[Glu,Tyr]4:1), in the presence of Mg^{2+} , Mn^{2+} and ^{32}P -labeled ATP. The reaction is terminated by "spotting" a sample onto a filter paper disc, followed by immersion in acid to precipitate the radiolabeled product. The filter papers are then washed extensively to remove unincorporated radiolabel and the radioactivity is counted. While sensitive, this method is labor-intensive, generates hazardous radioactive waste, and depends on a radioisotope of short half-life. It is particularly unsuitable when kinase assays are only performed on an infrequent basis. The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** uses a horseradish peroxidase coupled anti-phosphotyrosine monoclonal antibody as a reporter molecule in a 96-wells ELISA format. This assay provides a non-isotopic, sensitive and specific method to detect kinase activity of recombinant FGFR2 catalytic domain.



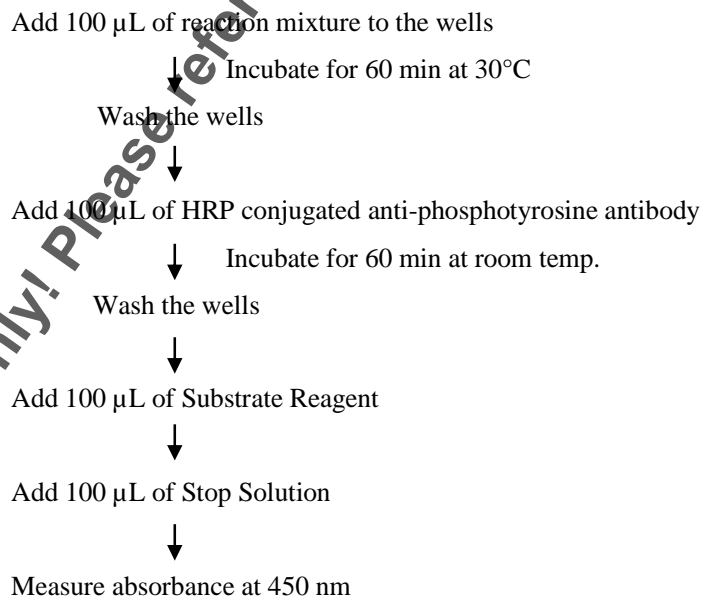
Principle of the Assay

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** is a single-site, semi-quantitative immunoassay for kinase activity of recombinant catalytic domain of FGFR2. Plates are pre-coated with a newly designed “Tyrosine kinase-substrate-1”, which can be easily phosphorylated by recombinant catalytic domain of FGFR2. The detector antibody is PY-39, an antibody that specifically detects the phosphotyrosine residue on “Tyrosine kinase-substrate-1”.

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** might be used to follow the kinetics of recombinant catalytic domain of FGFR2 as well as screening FGFR2 inhibitor or activator. To perform the test, the recombinant catalytic domain of FGFR2 is diluted in Kinase Buffer, pipetted into the wells and allowed to phosphorylate “Tyrosine kinase-substrate-1” in the presence of Mg^{2+} , Mn^{2+} and ATP. The amount of phosphorylated the recombinant “Tyrosine kinase-substrate-1” is measured by binding it with a horseradish peroxidase conjugate of PY-39, a anti-phosphotyrosine monoclonal antibody, which then catalyzes the conversion of the chromogenic substrate tetra-methylbenzidine (TMB) from a colorless solution to a blue solution (or yellow after the addition of stopping reagent). The color is quantified by spectrophotometry and reflects the relative amount of FGFR2 kinase activity in the sample. For kinetic analysis, the recombinant catalytic domain of FGFR2 is added to the wells in a similar fashion and at varying times the reaction is stopped by the addition of a chelator, sodium ethylenediaminetetraacetate (EDTA) and the amount of phosphorylated substrate determined as before.

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** is designed to determine non-isotopic kinetic analysis of the FGFR2 catalytic domain kinase activity. Careful attention to operation and the assay protocol will provide the investigator with a reliable tool for the evaluation of inhibitor or activator of FGFR2 kinase.

Summary of Procedure





Materials Provided

All samples and standards should be assayed in duplicate. The following components are supplied and are sufficient for the one 96-wells microtiter plate kit.

Microplate: One microplate supplied ready to use, with 96 wells (12 strips of 8-wells) in a foil, zip-lock bag with a desiccant pack. Wells are coated with recombinant "Tyrosine kinase-substrate-1".

10X Wash Buffer: One bottle containing 100 mL of 10X buffer containing 2 % Tween[®]-20

Kinase Buffer: One bottle containing 20 mL of 1X buffer used for Kinase Reaction Buffer and sample dilution.

20X ATP: One vial of lyophilized ATP Na₂ salt.

20X DTT: One vial of lyophilized dithiothreitol.

HRP conjugated Detection Antibody: One bottle containing 12 mL of HRP (horseradish peroxidase) conjugated anti-phosphotyrosine monoclonal antibody (PY-39).

Substrate Reagent: One bottle containing 20 mL of the chromogenic substrate, tetra-methylbenzidine (TMB). Ready to use.

Stop Solution: One bottle containing 20 mL of 1 N H₂SO₄. Ready to use.

Materials Required but not Provided

FGFR2 Positive Control: One vial contains 100 units recombinant catalytic domain of FGFR2. The Positive Control should be added to the first well at 1 unit/well. For instance, in the case of 100 units/100 μ L FGFR2 Positive Control, diluted Positive Control 1:10 with Kinase Buffer, use 10 μ L for 1 assay. The Positive Control is separately delivered with a dry ice from other kit components. The Positive Control should be stored in aliquots at **below -70°C**. Avoid repeated freeze and thaw.

- **10X Staurosporine (100 μ M):** Staurosporine is available from Sigma, Cat#. S-4400. 10 mM stock solution (DMSO) diluted 1:100 in Kinase Buffer.
- **10X Quercetin dihydrate:** 500 μ M Quercetin dihydrate (Sigma Cat# Q-0125: make 10 mM DMSO solution and diluted 1:20 in water)
- **Pipettors:** 2-20 μ L, 20-200 μ L and 200-1000 μ L precision pipettors with disposable tips.
- **Precision repeating pipettor**
- **Wash bottle or multichannel dispenser** for plate washing.
- **Microcentrifuge and tubes** for sample preparation.
- **Vortex mixer**
- **Plate reader** capable of measuring absorbance in 96-well plates at dual wavelengths of 450 nm/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. The plate can also be read at a single wavelength of 450 nm, which will give a somewhat higher reading.
- **Reagent reservoirs**
- **Deionized water of the highest quality**



Precautions and Recommendations

- Store the ATP at -20°C in aliquots. Store all other components at 4°C. Do not expose reagents to excessive light. Avoid freeze/thaw cycles.
- Allow all the components to come to room temperature before use.
- Do not use kit components beyond the indicated kit expiration date.
- Use only the microtiter wells provided with the kit.
- Rinse all detergent residue from glassware.
- Use deionized water of the highest quality.
- Do not mix reagents from different kits.
- The buffers and reagents in this kit may contain preservatives or other chemicals. Care should be taken to avoid direct contact with these reagents.
- Do not mouth pipet or ingest any of the reagents.
- Do not smoke, eat, or drink when performing the assay or in areas where samples or reagents are handled.
- Dispose of tetra-methylbenzidine (TMB) containing solutions in compliance with local regulations.
- Avoid contact with Substrate Solution which contains hydrogen peroxide.
- Avoid contact with Stop Solution which contains Sulfuric Acid.
- In case of contact with the Stop Solution and the Substrate Solution, wash skin thoroughly with water and seek medical attention, when necessary.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**
- **CAUTION: Sulfuric Acid is a strong acid. Wear disposable gloves and eye protection when handling Stop Solution.**



Detailed Protocol

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** is provided with removable strips of wells so the assay can be carried out on separate occasions using only the number of strips required for the particular determination. Due to technical reason, this kit is adjusted to measure kinase activity of the recombinant catalytic domain of FGFR2 (CycLex FGFR2 Positive Control: not provided, Cat# CY-E1082), which should be used in all assays. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

Preparation of Working Solution

1. Prepare a working solution of **Wash Buffer** by adding 100 mL of the **10X Wash Buffer** (provided) to 900 mL of ddH₂O. Mix well. Store at 4°C for two weeks or -20°C for long-term storage.
2. Prepare **20X ATP Solution** by adding **1.6 mL** of ddH₂O to the vial of **20X ATP** (provided, lyophilized). Mix gently until dissolved. The final concentration of the **20X ATP Solution** should be **1.25 mM**. Store the solution in small aliquots (e.g. 100 µL) at -20°C.
3. Prepare **20X DTT Solution** by adding **1.25 mL** of ddH₂O to the vial of **20X DTT** (provided, lyophilized). Mix gently until dissolved. The final concentration of the **20X DTT Solution** should be **40 mM**. Store the solution in small aliquots (e.g. 200 µL) at -20°C.
4. Prepare **Kinase Reaction Buffer** by mixing following reagents.

	96 assays	10 assays	1 assay
Kinase Buffer (provided)	9.0 mL	900 µL	90 µL
20X DTT Solution	0.5 mL	50 µL	5 µL
20X ATP Solution	0.5 mL	50 µL	5 µL
Total	10 mL	1000 µL	100 µL

You will need 80-90 µL of Kinase Reaction Buffer per assay well. Mix well. Discard any unused Kinase Reaction Buffer after use.

Standard Assay

1. Remove the appropriate number of microtiter wells from the foil pouch and place them into the well holder. Return any unused wells to the foil pouch, refold, seal with tape and store at 4°C.
2. Prepare the Kinase Assay Buffer containing test chemicals and tyrosine kinase inhibitor (see page 8). All assays should be done in duplicate.
3. Add **10 µL** of 0.1 unit/µL FGFR2 Positive Control or serial dilution of FGFR2 Positive Control to the wells of the assay plate on ice.
4. Begin the kinase reaction by addition of **90 µL Kinase Reaction buffer** per well, cover with plate sealer, and incubate **at 30°C for 60 minutes**.



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5. Wash wells five times with Wash Buffer making sure each well is filled completely. Remove residual Wash Buffer by gentle tapping or aspiration.
6. Pipette **100 μ L HRP conjugated Detection Antibody** into each well, cover with a plate sealer and incubate **at room temperature (ca.25°C) for 60 minutes**. Discard any unused conjugate.
7. Wash wells five times with Wash Buffer making sure each well is filled completely. Remove residual Wash Buffer by gentle tapping or aspiration.
8. Add **100 μ L of Substrate Reagent** to each well and incubate **at room temperature (ca.25°C) for 5–15 minutes**.
9. Add **100 μ L of Stop Solution** to each well in the same order as the previously added Substrate Reagent.
10. Measure absorbance in each well using a spectrophotometric plate reader at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. Read the plate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop Solution.

Note-1: Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

Note-2: Reliable signals are obtained when either O.D. values do not exceed 0.25 units for the blank (no enzyme control), or 2.5 units for the FGFR2 Positive Control.

Note-3: If the microplate reader is not capable of reading absorbance greater than the absorbance of the FGFR2 Positive Control, perform a second reading at 405 nm. A new O.D. values, measured at 405 nm, is used to determine FGFR2 activity of off-scale samples. The readings at 405 nm should not replace the on-scale readings at 450 nm.

Kinetic Assay

1. Remove the appropriate number of microtiter wells from the foil pouch and place them into the well holder. Return any unused wells to the foil pouch, refold, seal with tape and store at 4°C.
2. Prepare all samples (diluted with Kinase Buffer as needed). All samples should be assayed in duplicate.
3. Add **10 μ L** of 0.1 unit/ μ L FGFR2 Positive Control or serial dilution of FGFR2 Positive Control to the wells of the assay plate on ice.
4. Begin kinase reaction by addition of **90 μ L Kinase Reaction Buffer** in duplicate per well in timed intervals (suggested interval is 5 minutes but should be individually determined for each system). After the final addition, incubate **at 30°C for 20 minutes**.
5. Stop the reaction by flicking out the contents. (Alternatively, the reaction may be terminated by the addition of 150 μ L 0.1 M Na EDTA, pH 8.0 to each well).
6. Wash wells five times with Wash Buffer making sure each well is filled completely. Remove



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residual Wash Buffer by gentle tapping or aspiration.

- Pipette **100 μ L HRP conjugated Detection Antibody** into each well, cover with a plate sealer and incubate **at room temperature (ca.25°C) for 60 minutes**. Discard any unused conjugate.
- Wash wells five times with Wash Buffer making sure each well is filled completely. Remove residual Wash Buffer by gentle tapping or aspiration.
- Add **100 μ L of Substrate Reagent** to each well and incubate **at room temperature (ca.25°C) for 5-15 minutes**.
- add **100 μ L of Stop Solution** to each well in the same order as the previously added Substrate Reagent.
- Measure absorbance in each well using a spectrophotometric plate reader at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. Read the plate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop Solution.

Recommendations

Special considerations when screening activators and inhibitors

In order to estimate the inhibitory effect on FGFR2 activity in the test chemicals correctly, it is necessary to conduct the control experiment of "Solvent control" at least once for every experiment and "Inhibitor control" at least once for the first experiment, in addition to "Test sample", as indicated in the following table. When test chemicals cause an inhibitory effect on FGFR2 activity, the level of A450 is weakened as compared with "Solvent control". The high level of A450 is not observed in "Inhibitor control" (usually A450<0.4).

Assay reagents	Test sample	Solvent control	Inhibitor control
Kinase Reaction Buffer	80 μL	80 μL	80 μL
10X Inhibitor or equivalent	10 μL	-	-
Solvent for Inhibitor	-	10 μL	-
10X Staurosporine (100 μM)*	-	-	10 μL
FGFR2 Positive Control (0.1 unit/μL)**	10 μL	10 μL	10 μL

* 10X Staurosporine (100 μ M): See page 4, section "Materials Required but not Provided"

** FGFR2 Positive Control (0.1 unit/ μ L)**: See page 4, section "Materials Required Provided"

- Following the above table, add the Reagents to each well of the microplate. Finally, initiate reaction by adding 10 μ L of "Diluted CycLex FGFR2 Positive Control" to each well and mixing thoroughly at room temperature. Cover with plate sealer. Incubate at 30°C for 60 minutes.
- Follow the **Standard Assay**, steps 5-10, page 6-7.



Evaluation of Results

1. Average the absorbance values for the FGFR2 sample duplicates (Positive Control) and all experimental sample duplicate values (when applicable). When the FGFR2 Positive Control (1 unit/assay) is included as an internal control for the phosphorylation reaction, the absorbance value should be greater than 1.0 with a background less than 0.2.
2. For kinetic analysis, on graph paper, plot the mean absorbance values for each of the time points on the Y-axis versus the time of each reaction (minutes) on the X-axis.

Assay Characteristics

The CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** has been shown to detect the kinase activity of recombinant catalytic domain of FGFR2. The assay shows good linearity of sample response.

Troubleshooting

1. The FGFR2 Positive Control should be run in duplicate, using the protocol described in the **Detailed Protocol**. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. The reaction curve is nearly a straight line if the kinetics of the assay is of the first order. Variations in the protocol can lead to non-linearity of the curve, as can assay kinetics that are other than first order. For a non-linear curve, point to point or quadratic curve fit methods should be used.
3. Poor duplicates, accompanied by elevated values for wells containing no sample, indicate insufficient washing. If all instructions in the **Detailed Protocol** were followed accurately, such results indicate a need for washer maintenance.
4. Overall low signal may indicate that desiccation of the plate has occurred between the final wash and addition of Substrate Reagent. Do not allow the plate to dry out. Add Substrate Reagent immediately after wash.

Reagent Stability

All of the reagents included in the CycLex Research Product **CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit** have been tested for stability. Reagents should not be used beyond the stated expiration date. Upon receipt, kit reagents should be stored at 4°C. Coated assay plates should be stored in the original foil bag sealed by the zip lock and containing a desiccant pack.

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Example of Test Results

Fig.1 Dose dependency of recombinant FGFR2 catalytic domain enzyme reaction

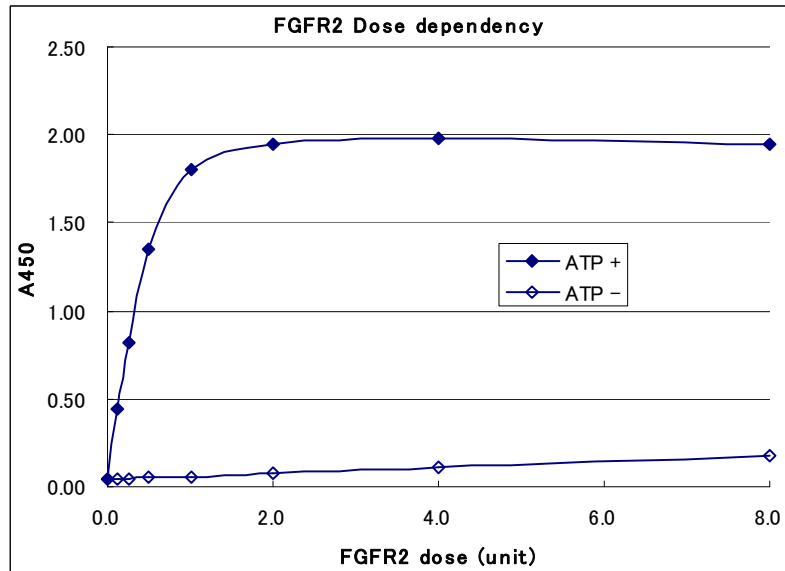
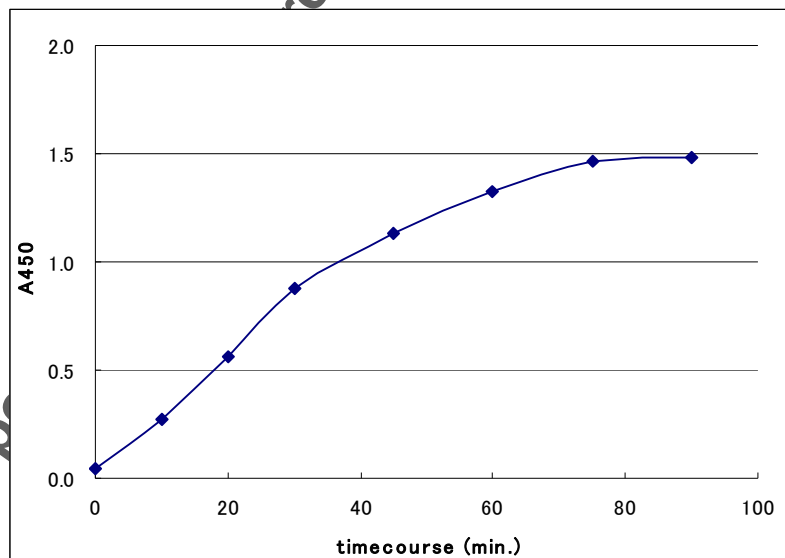


Fig.2 Time course of recombinant FGFR2 catalytic domain enzyme reaction





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Fig.3-1 Dose dependency of ATP (of recombinant FGFR2 catalytic domain)

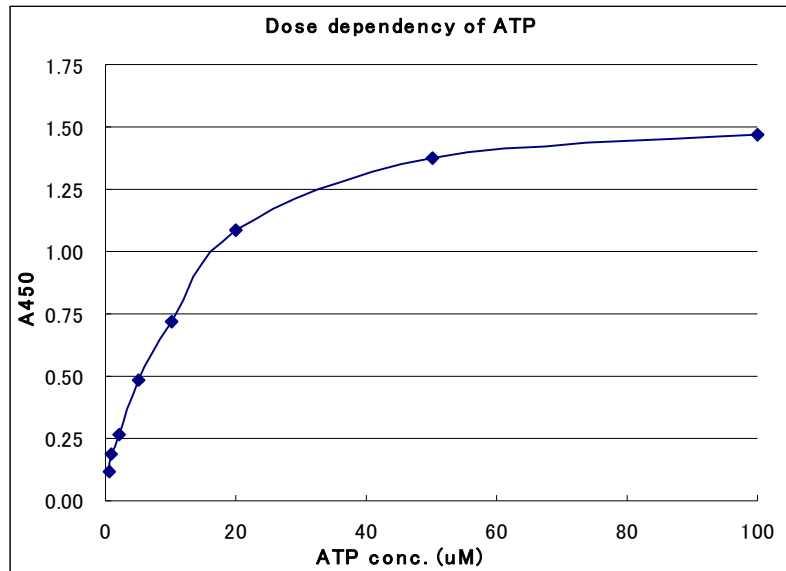
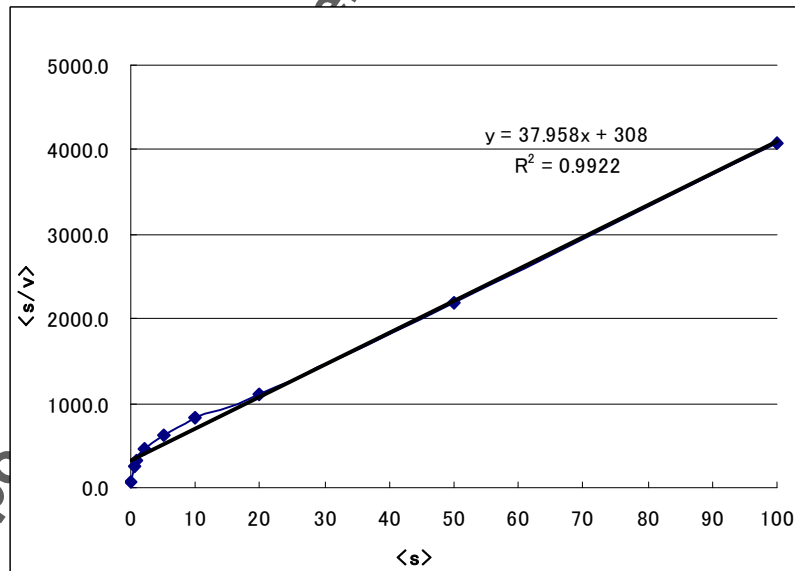


Fig.3-2 Km for ATP (of recombinant FGFR2 catalytic domain)





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Fig.4-1 Effect of broad-spectrum kinase inhibitor staurosporine on activity of recombinant FGFR2 catalytic domain enzyme reaction

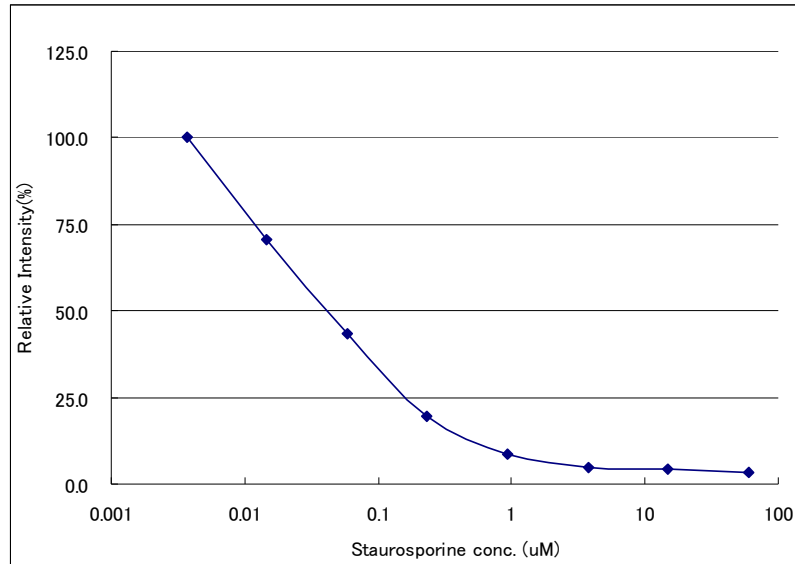
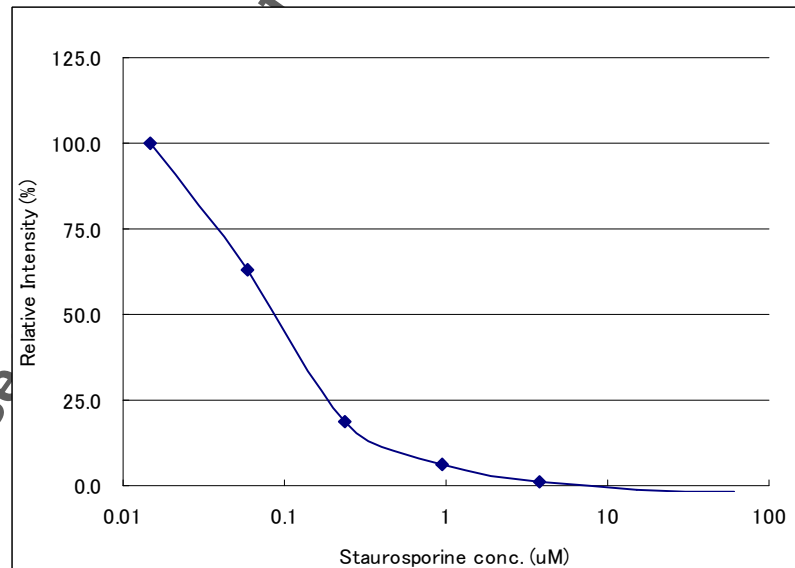


Fig.4-2 Effect of broad-spectrum kinase inhibitor staurosporine on activity of recombinant FGFR2 catalytic domain enzyme reaction using radioisotope gamma-³²P-ATP





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Related Products

Tyrosine Kinase Assay Kit

- * CycLex Wee1 Kinase Assay/Inhibitor Screening Kit: Cat# CY-1172
- * CycLex Met Kinase Assay/Inhibitor Screening Kit: Cat# CY-1080
- * CycLex Pyk2 Kinase Assay/Inhibitor Screening Kit: Cat# CY-1081
- * CycLex FGFR2 Kinase Assay/Inhibitor Screening Kit: Cat# CY-1082
- * CycLex c-Src Kinase Assay/Inhibitor Screening Kit: Cat# CY-1083
- * CycLex p56Lck Kinase Assay/Inhibitor Screening Kit: Cat# CY-1084
- * CycLex TrkA Kinase Assay/Inhibitor Screening Kit: Cat# CY-1091
- * CycLex EphA2 Kinase Assay/Inhibitor Screening Kit: Cat# CY-1092

Tyrosine Kinase Positive Control

- * Wee1 Positive Control: Cat# CY-E1172
- * Met Positive Control: Cat# CY-E1080
- * Pyk2 Positive Control: Cat# CY-E1081
- * FGFR2 Positive Control: Cat# CY-E1082
- * c-Src Positive Control: Cat# CY-E1083
- * p56Lck Positive Control: Cat# CY-E1084
- * TrkA Positive Control: Cat# CY-E1091
- * EphA2 Positive Control: Cat# CY-E1092

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